

## **Molecular dynamics simulations of charged and neutral lipid bilayers: treatment of electrostatic interactions<sup>★✉</sup>**

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Received: 30 May, 2003; revised: 24 July, 2003; accepted: 01 September, 2003

**Key words:** particle-mesh Ewald, cut-off distance, membrane hydration, chain order, lateral diffusion, phospholipid

Molecular dynamics (MD) simulations complement experimental methods in studies of the structure and dynamics of lipid bilayers. The choice of algorithms employed in this computational method represents a trade-off between the accuracy and real calculation time. The largest portion of the simulation time is devoted to calculation of long-range electrostatic interactions. To speed-up evaluation of these interactions, various approximations have been used. The most common ones are the truncation of long-range interactions with the use of cut-offs, and the particle-mesh Ewald (PME) method. In this study, several multi-nanosecond cut-off and PME simulations were performed to establish the influence of the simulation protocol on the bilayer properties. Two bilayers were used. One consisted of neutral phosphatidylcholine molecules. The other was a mixed lipid bilayer consisting of neutral phosphatidylethanolamine and negatively charged phosphatidylglycerol molecules. The study shows that the cut-off simulation of a bilayer containing charge molecules generates artefacts; in particular the mobility and order of the charged molecules are vastly different from those

<sup>★</sup>Presented at the XXX Winter School of Faculty of Biotechnology, Jagiellonian University, Kościelisko, Poland, 28th February–4th March, 2003.

<sup>✉</sup>This work was supported in part by grant 6P04A03121 from the State Committee for Scientific Research (KBN, Poland) and by the European Union (contract no. BIER ICA1-CT-2000-70012). All calculations were performed at the academic Computer Centre Cyfronet, Kraków, Poland, grants number KBN/sgi\_origin\_200/004/2000 and KBN/sgi\_origin\_2000/062/1999.

<sup>\*</sup>T.R. holds a fellowship award from the Foundation for Polish Science in 2002/2003. K.M. acknowledges a fellowship award from the Foundation for Polish Science in 2001/2002.

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**Abbreviations:** MD, molecular dynamics; OPLS, optimised potential for liquid simulations; PME, particle-mesh Ewald; POPC, 1-palmitoyl-2-oleoyl-phosphatidylcholine; POPE, 1-palmitoyl-2-oleoyl-phosphatidylethanolamine; POPG, 1-palmitoyl-2-oleoyl-phosphatidylglycerol; TIP, transferable intermolecular potential.

determined experimentally. In the PME simulation, the bilayer properties are in general agreement with experimental data. The cut-off simulation of bilayers containing only uncharged molecules does not generate artefacts, nevertheless, the PME simulation gives generally better agreement with experimental data.

Molecular dynamics (MD) simulation is a valuable method to study molecular systems of biological relevance. The method is particularly well suited to investigating the dynamical structure of biomembranes because it is characterised by atomic resolution and allows observation of the processes on the time scale of  $10^{-8}$ – $10^{-7}$  s with a time resolution of a few femtoseconds. To carry out MD simulations of a molecular system, relevant atom–atom interactions have to be defined. Of all inter-atomic interactions that are commonly included in the potential function of the system, the electrostatic ones have the largest contribution at longer distances. A correct treatment of long-range electrostatic interactions is essential for obtaining meaningful simulation results. Since full evaluation of Coulombic interactions requires enormous computational resources, various approximations have been used. The most common are (a) truncation of long-range interactions with the use of cut-offs and (b) particle-mesh Ewald (PME) summation (Essmann *et al.*, 1995). The theoretical grounds of the two methods is discussed in depth in Sagui & Daren (1999). The use of cut-offs has been shown to lead to several artefacts in simulations of peptides (Schreiber & Steinhauser, 1992) and nucleic acids (York *et al.*, 1995). In the case of lipid bilayers, cut-off simulations reproduce experimental data sufficiently well (e.g., Pasenkiewicz-Gierula *et al.*, 1999; Tieleman & Berendsen, 1998). However, this observation is based on the studies of bilayers consisting of phospholipids with zero net charge.

In this paper, cut-off and PME MD simulations of bilayers formed of uncharged and charged phospholipids are compared. The study shows that MD simulation of a bilayer containing charged molecules gives reasonable results only when electrostatic interactions are calculated using the PME summa-

tion method. Application of the PME method to uncharged bilayers does not significantly improve the results. The latter conclusion is at variance with the conclusion of Patra *et al.* (2003) who claim that bilayers made of neutral phospholipids generated in the cut-off and Ewald MD simulations were different. They used, however, an unjustified approach for the evaluation of electrostatic interactions.

## METHOD

### Simulation systems

In this MD simulation study two phospholipid bilayers were used. One contained both neutral and charged phospholipids (POPE-POPG bilayer); the other contained only neutral phospholipids (POPC bilayer). The POPE-POPG bilayer consisted of 54 1-palmitoyl-2-oleoyl-phosphatidylethanolamine (POPE), 18 1-palmitoyl-2-oleoyl-phosphatidylglycerol (POPG) (25 mol% POPG), and 1955 water molecules, and of 18 sodium ions ( $\text{Na}^+$ ). The cations were added to neutralise the net charge of  $-1.0$  in units of electronic charge ( $e$ ) on each POPG molecule. Details concerning the membrane construction are given in (Murzyn & Pasenkiewicz-Gierula, 1999). An initial 11-ns simulation of the POPE-POPG bilayer was carried out using AMBER 4.0 (Pearlman *et al.*, 1991). In this part of the simulation, a residue based cut-off distance of 12 Å was used to evaluate the nonbonded interactions. The last 6-ns fragment of the generated trajectory was analysed. The simulation was continued for another 14 ns using AMBER 5.0 (Case *et al.*, 1997). In this part of the simulation the electrostatic interactions were evaluated using the PME summation method. The last 10-ns fragment of this trajectory was analysed.

The POPC bilayer consisted of 72 1-palmitoyl-2-oleoyl-phosphatidylcholine (POPC) and 1955 water molecules. Details concerning the membrane construction are given in (Murzyn *et al.*, 2001). An initial 11-ns simulation of the bilayer was carried out using AMBER 4.0 and a 12 Å cut-off. The simulation was continued for 6 ns using AMBER 5.0 and

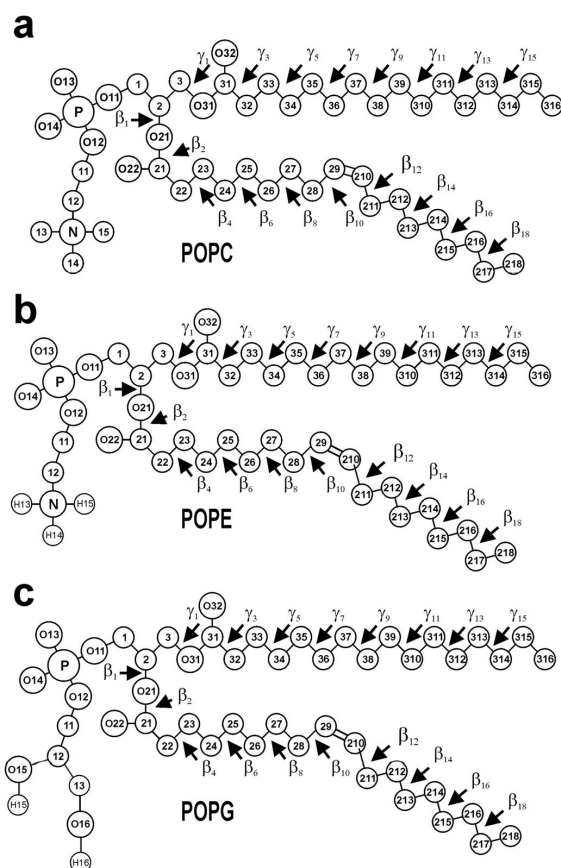
### Simulation parameters

For POPC, POPE, and POPG, optimised potentials for liquid simulations (OPLS) parameters (Jorgensen & Tirado-Rives, 1988), and for water, transferable intermolecular potential three-site model (TIP3P) parameters (Jorgensen *et al.*, 1983) were used. The united-atom approximation was applied to the CH, CH<sub>2</sub>, and CH<sub>3</sub> groups of the phospholipids. The numerical values for the atomic charges of POPE and POPG are given in Murzyn and Pasenkiewicz-Gierula (1999) and of POPC follow those in Charifson *et al.* (1990). Other details concerning the POPE-POPG bilayer can be found in Murzyn & Pasenkiewicz-Gierula (1999) and the POPC bilayer in Murzyn *et al.* (2001).

### Simulation conditions

Three-dimensional periodic boundary conditions with the usual minimum image convention were used. The SHAKE algorithm (Ryckaert *et al.*, 1977) was used to preserve the bond lengths of the OH and NH<sub>3</sub> groups of water, POPE, and POPG molecules, and the time step was set at 2 fs (Egberts *et al.*, 1994). The list of nonbonded pairs was updated every 25 steps. Restraints of a flat-bottom harmonic potential as defined and implemented in the AMBER package (Case *et al.*, 1997) were imposed on the double bonds to prevent *cis-trans* isomerisation. To evaluate the nonbonded interactions at first a residue base 12 Å cut-off was used (cut-off simulation) and then a PME summation (Essmann *et al.*, 1995) with a real cut-off of 12 Å,  $\beta$  spline interpolation order of 5, and direct sum tolerance of  $10^{-6}$  was used (PME simulation).

The MD simulations were carried out at constant pressure (1 atm) and at the temperature of 310 K (37°C), which is above the main phase transition temperature for POPC (−5°C) (Seelig & Waespe-Sarčević, 1978), POPG (−4°C) (LIPIDAT), and POPE (26.1°C) (Huang & Li, 1999) bilayers. The temperatures of the solute and solvent were controlled



**Figure 1.** Molecular structure with numbering of atoms and torsion angles of POPC (a), POPE (b), and POPG (c) molecules (chemical symbol for carbon atoms, C, and their attached H atoms have been omitted).

a PME summation (Essmann *et al.*, 1995) with the same parameters as in the case of the POPE-POPG bilayer (see below). Last 4-ns fragments of both trajectories (cut-off and PME) were used for analysis. Figure 1 shows the structure and numbering of atoms and torsion angles in POPC, POPE, and POPG molecules.

independently. Both the temperature and pressure of the system were controlled by the Berendsen method (Berendsen *et al.*, 1984). The relaxation times for temperatures and pressure were set at 0.4 and 0.6 ps, respectively. The applied pressure was controlled anisotropically, each direction being treated independently with the trace of the pressure tensor kept constant at 1 atm.

## RESULTS

The average values given below are ensemble and time averages obtained from the block averaging procedure. Errors in the derived average values are standard error estimates.

### Surface area per phospholipid and bilayer thickness

A two-dimensional Voronoi tessellation method (Shinoda & Okazaki, 1998) allows one to compute the surface area of a single molecule in the bilayer made of lipids of a single type (cf. Murzyn *et al.*, 2001). In the case of a mixed lipid bilayer, the method is less precise, nevertheless it was applied to estimate the surface area of each POPE and POPG molecule in the POPE-POPG bilayer. From the estimated single molecular areas, the average values for POPE and POPG in both cut-off and Ewald simulations were obtained (Table 1). The average surface area per POPC in the POPC bilayer was obtained in a straightforward way by dividing the total surface area of the bilayer by 36, which is the number of molecules in each leaflet of the bilayer. The values for both cut-off and Ewald simulations are given in Table 1.

The bilayer thickness was estimated from the distance between average positions of the phosphorus atoms in the two leaflets of a bilayer (P-P distance). The values for both cut-off and Ewald simulations are given in Table 1.

**Table 1. Bulk properties of POPC and POPE-POPG bilayers in Ewald and cut-off simulations.**

Surface area per phospholipid and bilayer thickness (P-P distance) are listed with standard error estimates.

		Ewald	Cut-off
POPC	Surface ( $\text{\AA}^2$ )	$63.5 \pm 0.3$	$64.3 \pm 0.3$
	P-P distance ( $\text{\AA}$ )	$35.5 \pm 0.1$	$35.5 \pm 0.1$
POPE	Surface ( $\text{\AA}^2$ )	$61.0 \pm 0.4$	$58.3 \pm 0.4$
	P-P distance ( $\text{\AA}$ )	$36.0 \pm 0.1$	$40.0 \pm 0.1$
POPG	Surface ( $\text{\AA}^2$ )	$62.8 \pm 0.4$	$63.9 \pm 0.4$
	P-P distance ( $\text{\AA}$ )	$36.0 \pm 0.1$	$38.7 \pm 0.1$

Both the surface area per phospholipid and the bilayer thickness are more affected by the simulation protocol in the case of the POPE-POPG bilayer than the POPC bilayer, but the effect is not very large.

### Bilayer/water interface

Intermolecular interactions that take place at the bilayer/water interface include hydrogen bonding between water molecules and polar groups of POPC, POPE, and POPG; direct H-bonding and charge pairing among POPC, POPE, and POPG headgroups; and clathrating of water molecules around the choline group of POPC. A water molecule that is simultaneously H-bonded to two phospholipid molecules forms a water bridge between them, whereas a charge pair is formed between a negatively charged oxygen atom of one POPC and a positively charged choline moiety of a neighbouring POPC. In the following analyses, the same geometrical definitions of H-bonding, water bridging, and charge pairing as in our previous papers (Pasenkiwicz-Gierula *et al.*, 1999; Murzyn *et al.*, 2001) were used. In addition to these short distance



intermolecular interactions, in the POPE-POPG bilayer we define charge pairs between  $\text{Na}^+$  ions and oxygen atoms of phosphate, carbonyl, or glycerol groups (cf. Fig. 1), when the ion and the oxygen atom are no more than 3.0 Å apart.

### POPC bilayer

Numbers of water molecules that interact with POPC as well as of POPC-POPC charge pairs for the two simulation protocols are given in Table 2. The numbers are practically

**Table 2. Intermolecular interactions at the bilayer/water interface for Ewald and cut-off simulations of the POPC bilayer.**

Listed are average numbers of water molecules H-bonded to POPC and in a clathrate around the POPC choline group (choline water), of bound water molecules (H-bonded and choline water), of water bridges, and of charge pairs per POPC. Errors are standard error estimates.

No./PC	Ewald	Cut-off
H-bonded water	$4.9 \pm 0.1$	$5.0 \pm 0.1$
Choline water	$6.4 \pm 0.1$	$6.6 \pm 0.2$
Bound water	$11.3 \pm 0.1$	$11.6 \pm 0.2$
Water bridges	$0.6 \pm 0.1$	$0.5 \pm 0.1$
Charge pairs	$1.7 \pm 0.1$	$1.7 \pm 0.1$

unaffected by the simulation protocol, as differences between the corresponding numbers for the cut-off and Ewald simulations are within estimated errors.

### POPE-POPG bilayer

Numbers of water molecules that interact with POPE and POPG, of direct H-bonds between phospholipids, and of charge pairs between phospholipids and  $\text{Na}^+$  for the cut-off and Ewald simulations are given in Table 3. In contrast to the POPC bilayer, the numbers of

intermolecular interactions at the bilayer/water interface of the POPE-POPG bilayer strongly depend on the simulation protocol. The number of water molecules H-bonded to a phospholipid is significantly higher, whereas the number of the lipid-lipid H-bonds is smaller in the cut-off simulation. A considerable difference in the number of  $\text{Na}^+$ -lipid charge pairs for the two protocols is noticeable. In the cut-off simulation,  $\text{Na}^+$  practically do not interact with POPE but do very frequently interact with POPG. On average, 50% of all  $\text{Na}^+$  ions are bound to POPG at any instant. In the Ewald simulation, only 30% of all  $\text{Na}^+$  ions are bound to lipids, 20% to POPG and 10% to POPE (Table 3).

### Bilayer core

The molecular order parameter for the  $n$ th segment of an alkyl chain,  $S_{\text{mol}}$ , is defined through (Hubbell & McConnell, 1971)

$$S_{\text{mol}} = \frac{1}{2} \langle 3 \cos^2 \theta_n - 1 \rangle$$

$\theta_n$  is the instantaneous angle between the  $n$ th segmental vector, i.e., the  $(C_{n-1}, C_{n+1})$  vector linking the  $n - 1$  and  $n + 1$  carbon atoms in the alkyl chain and the bilayer normal;  $\langle \rangle$  denotes the ensemble and time average.  $S_{\text{mol}}$  profiles along the  $\beta$ -chain of POPC, POPE, and POPG molecules are shown in Fig. 2. Details concerning calculation of conformation-related quantities are given in Róg and Pasenkiewicz-Gierula (2001) and Murzyn *et al.* (1999). Mean values of the order parameter for the  $\beta$ -chain for the cut-off and Ewald simulations are given in Table 4. The order of the  $\beta$ -chain is, in general, higher for the PME than the cut-off simulation. A particularly large difference is observed for the  $\beta$ -chain of POPG (Fig. 2 c, and Table 4).

Other parameters characterising the hydrocarbon chain region of the bilayer, like the  $\beta$ -chain tilt angle (for definition see Róg *et al.*, 2003\*) and the number of *gauche* rotamers in

**Table 3. Intermolecular interactions at the bilayer/water interface for Ewald and cut-off simulations of the POPE-POPG bilayer.**

Listed are average numbers of water molecules H-bonded to a lipid molecule, of direct lipid-lipid H-bonds, of water bridges, and of  $\text{Na}^+$ -lipid charge pairs per POPE and POPG molecule. Errors are standard error estimates.

No./PC	Lipid	Ewald	Cut-off
H-bonded water	POPE	5.8± 0.1	7.3± 0.1
	POPG	5.1± 0.1	8.3± 0.1
Direct lipid-lipid H-bonds	POPE	1.1± 0.1	0.6± 0.1
	POPG	1.3± 0.1	0.9± 0.1
Water bridges	POPE	1.6± 0.1	1.9± 0.1
	POPG	2.0± 0.1	2.6± 0.1
$\text{Na}^+$ -lipid charge pairs	POPE	0.1± 0.1	0.04± 0.01
	POPG	0.2± 0.1	0.5± 0.1

the  $\beta$ -chain are also given in Table 4. These parameters are affected only mildly by the simulation protocol.

#### Lateral diffusion of phospholipids

Diffusion coefficients for the lateral self-diffusion of POPC, POPE, and POPG molecules in the bilayer plane ( $D$ ) were calculated from linear parts of the respective mean square dis-

placement and Ewald simulations are compared in Table 4. For POPC and POPE, the differences between the respective values of  $D$  for the two protocols are small. For POPG, the value of  $D$  in the cut-off simulation is 35 times smaller than in the Ewald simulation. A good illustration of the huge difference between the diffusion of POPG in the cut-off and Ewald simulations are the trajectories of the centre of mass of a selected POPG molecule displayed in

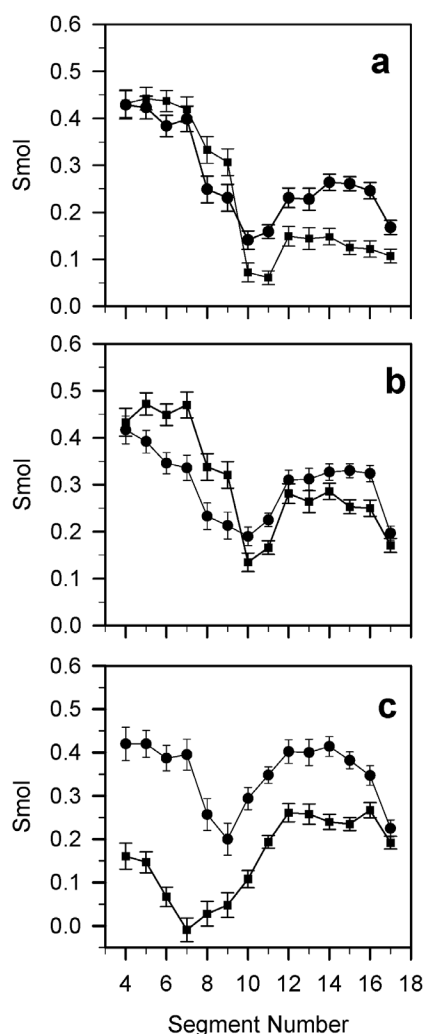
**Table 4. Average values of parameters for Ewald and cut-off simulations of POPC and POPE-POPG bilayers.**

Average values of molecular order parameter ( $S_{\text{mol}}$ ), tilt angle, and number of *gauche* conformations per chain, for the  $\beta$ -chain as well as the lateral diffusion coefficient ( $D$ ) for the centre of mass of POPC, POPE, and POPG are given with standard error estimates.

Bilayer	POPC		POPE		POPG	
	Ewald	Cut-off	Ewald	Cut-off	Ewald	Cut-off
$S_{\text{mol}}$	0.27 ± 0.01	0.24 ± 0.01	0.30 ± 0.01	0.30 ± 0.01	0.35 ± 0.01	0.16 ± 0.03
Tilt [°]	20.0 ± 0.5	25.0 ± 0.5	23.0 ± 0.5	26.0 ± 0.5	19.5 ± 0.5	21.0 ± 0.5
No. <i>gauche</i>	3.4 ± 0.1	3.5 ± 0.1	3.2 ± 0.1	2.9 ± 0.1	3.1 ± 0.1	3.3 ± 0.1
$D \cdot 10^{-7} \times [\text{cm}^2/\text{s}]$	4.5	4.0	3.5	3.3	3.5	0.1

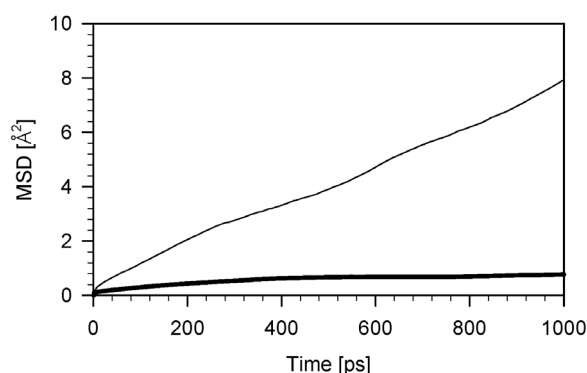
placement (MSD) curves for the centre of mass of all lipids of a given type in the membrane (Fig. 3). The coefficients for the cut-off

Fig. 4. In the cut-off simulation, the centre of mass of the POPG practically remains in the same position during a period of 4 ns, whereas



**Figure 2.** Profiles of the molecular order parameter  $S_{mol}$  along the  $\beta$ -chain in POPC (a), POPE (b), and POPG (c) in PME (●) and cut-off (■) simulations.

The error bars are standard errors.



**Figure 3.** Mean square displacement (MSD) curves for the centre of mass of POPG molecules in the plane of the POPE-POPG bilayer in PME (thin line) and cut-off (thick line) simulations.

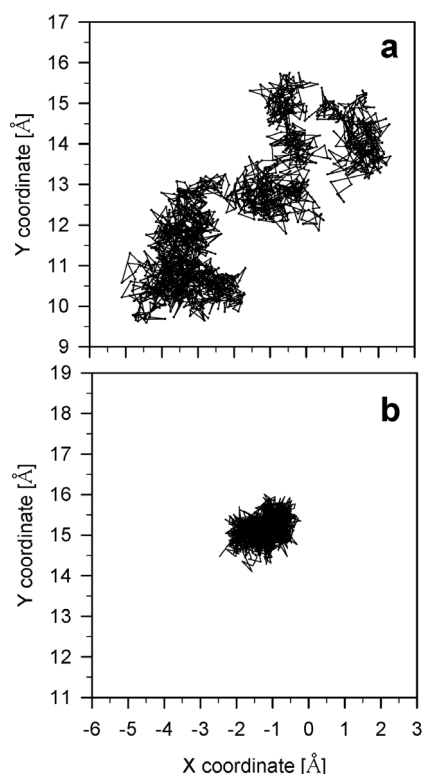
in the Ewald simulation, in this period it covers a large area.

## DISCUSSION

In this study, the effect of the MD simulation protocol on the properties of a bilayer made of uncharged and charged phospholipids has been evaluated. Two simulation protocols were applied. In one, long-range electrostatic interactions were truncated with the use of a residue-based 12  $\text{\AA}$  cut-off, in the other, the interactions were evaluated with the use of the PME summation method. Two bilayers were simulated and analysed. One consisted of neutral POPC molecules, the other consisted of neutral POPE and charged ( $-1.0 e$ ) POPG molecules.

The study clearly shows a strong effect of the simulation protocol on the behaviour of POPG but not POPE, although both POPGs and POPEs in the proportion 1:3 comprise one bilayer. The most striking result is the almost complete immobilisation of POPG molecules in the cut-off simulation as compared to POPE molecules in the same simulation and to their mobility in the PME simulation. In the cut-off simulation, the diffusion coefficient for POPG self-diffusion on the nanosecond time scale is over 30 times smaller than that of POPE and POPC, and than that of POPG in the PME simulation. The diffusion coefficients of PCs measured experimentally by neutron scattering (Tabony & Perly, 1991) and electric dipole relaxation (Haibel *et al.*, 1998) methods, which report events on a similar time scale as MD simulations, are similar to those of POPC and POPE obtained in both cut-off and PME simulations as well as to the value for POPG obtained in the PME simulation.

The simulation protocol also affects the molecular order parameter profile of POPG rather strongly but not POPE.  $S_{mol}$  profiles for hydrocarbon chains are available in the literature for phosphatidylcholines (PC) and phosphatidylethanolamines (PE) but not for



**Figure 4.** A sample 4-ns trajectory of the centre of mass of an arbitrary POPG molecule in the  $x,y$ -plane of the POPE-POPG bilayer in PME (a) and cut-off (b) simulations.

phosphatidylglycerols (PG). The  $S_{\text{mol}}$  profile of the POPG  $\beta$ -chain obtained in the cut-off simulation does not resemble the  $S_{\text{mol}}$  profiles derived experimentally for saturated (Touard *et al.*, 1999) or unsaturated (Seelig & Waespe-Sarčević, 1978) PC's or for PE's (Marsh *et al.*, 1983). However, that obtained in the PME simulation does resemble these profiles. The  $S_{\text{mol}}$  profiles for POPE obtained in both cut-off and PME simulations show a similar resemblance to experimental data.

An effect of the simulation protocol on the intermolecular interactions in the interfacial region of the POPE-POPG bilayer is also visible. In this region, both POPE and POPG are similarly affected. The cut-off simulation “over-produces” lipid–water H-bonds and “under-produces” lipid–lipid H-bonds. Consistently with the immobilisation of POPG in the cut-off simulation, the  $\text{Na}^+$  ions are predominantly in the bound state with POPG. In the PME simulation, the  $\text{Na}^+$  ions are more mo-

bile with fewer bound to the lipids; moreover, they interact with both POPE and POPG.

The effect of the simulation protocol on the POPC bilayer is much smaller than in the case of the POPE-POPG bilayer. Intermolecular interactions at the bilayer/water interface are practically unaffected. The order of the POPC alkyl chains better reproduces experimental data (Seelig & Waespe-Sarčević, 1978) in the PME simulation but the difference is not large. In a recent paper, results of 12 Å cut-off MD simulations of bilayers made of saturated, *trans*-, and *cis*-unsaturated lipids were compared with available experimental data (Róg *et al.*, 2003). The comparison indicated a good agreement between both sets of data.

The first and main conclusion of our study is that cut-off simulation of a bilayer containing, among others, molecules of non-zero net charge, generates artefacts. These artefacts concern the mobility and order of the charged molecules but not of the uncharged ones within the same bilayer. In the PME simulation, the bilayer properties are in general agreement with experimental data. Thus, whenever a bilayer system contains charged molecules, reasonable results of MD simulation can only be obtained when the long-range electrostatic interactions are evaluated using the PME summation method.

The second conclusion of our study is that cut-off simulation of a bilayer containing only uncharged molecules does not generate artefacts. The parameters that describe bilayer properties, derived from both cut-off and PME simulations are similar, although in some cases the results of the PME simulation better reproduce the relevant experimental data. The MD simulation studies of uncharged bilayers that we have undertaken so far provided qualitatively correct results even though neither cut-off nor PME simulations reproduce faithfully all relevant experimental results. To minimise the effects of methodological inaccuracies and other uncertainties, it is advisable always to conduct comparative MD simulation studies in which two or more



bilayer systems are simulated using the same protocol with one of the systems earlier having been compared successfully with appropriate experimental data. As the possible errors in parameters or methodology are most likely to be the same for all systems, such studies can be expected to give a faithful account of both similarities and differences among the systems.

In a recent paper, Patra *et al.* (2003) concluded that the properties of a bilayer made of neutral dipalmitoylphosphatidylcholine molecules generated in the cut-off and Ewald simulations were different. In those simulations, several parameters and definitions differed from those used by us. Particularly affecting the time development of the simulated system was their definition of a group in the group-based (our residue-based) cut-off simulation, where the sum of charges of all atoms comprising the group was far from zero, as well as the use of cut-off distances much larger than those for which OPLS parameters were derived. These unjustified choices caused significant discrepancies in the behaviour of their and our systems and resulted in opposite conclusions.

## REFERENCES

- Berendsen HJC, Postma JPM, van Gunsteren WF, DiNola A, Haak JR. (1984) Molecular dynamics with coupling to an external bath. *J Chem Phys.*; **81**: 3684–90.
- Case DA, Pearlman DA, Caldwell JW, Cheatham III TE, Ross WS, Simmerling C, Darden TA, Merz KM, Stanton RV, Cheng AL, Vincent JJ, Crowley M, Ferguson DM, Radmer RJ, Seibel GL, Singh UC, Weiner PK, Kollman PA. (1997) *AMBER 5.0*. University of California, San Francisco.
- Charifson PS, Hiskey RG, Pedersen LG. (1990) Construction and molecular modeling of phospholipid surfaces. *J Comp Chem.*; **11**: 1181–6.
- Egberts E, Marrik S-J, Berendsen HJC. (1994) Molecular dynamics simulation of phospholipid membrane. *Eur Biophys J.*; **22**: 423–36.
- Essmann U, Perera L, Berkowitz ML, Darden T, Lee H, Pedersen LG. (1995) A smooth particle mesh Ewald method. *J Chem Phys.*; **103**: 8577–93.
- Haibel A, Nimtz G, Pelster R, Jaggi R. (1998) Translational diffusion in phospholipid bilayer membranes. *Phys Rev E.*; **57**: 4838–41.
- Huang C, Li S. (1999) Calorimetric and molecular mechanics studies of the thermotropic phase behavior of membrane phospholipids. *Biochim Biophys Acta.*; **1422**: 273–307.
- Hubbell WL, McConnell HM. (1971) Molecular motion in spin-labeled phospholipids and membranes. *J Am Chem Soc.*; **93**: 314–26.
- Jorgensen WL, Tirado-Rives J. (1988) The OPLS potential functions for proteins: energy minimization for crystals of cyclic peptides and crambin. *J Am Chem Soc.*; **110**: 1657–66.
- Jorgensen WL, Chandrasekhar J, Madura JD, Impey R, Klein ML. (1983) Comparison of simple potential functions for simulating liquid water. *J Chem Phys.*; **79**: 926–35.
- LIPIDAT, <http://www.lipidat.chemistry.ohio-state.edu>
- Marsh D, Watts A, Smith ICP. (1983) Dynamic structure and phase behavior of dimyristoylphosphatidylethanolamine bilayers studied by deuterium nuclear magnetic resonance. *Biochemistry.*; **22**: 3023–6.
- Murzyn K, Pasenkiewicz-Gierula M. (1999) Construction and optimisation of a computer model for a bacterial membrane. *Acta Biochim Polon.*; **46**: 631–9.
- Murzyn K, Róg T, Pasenkiewicz-Gierula M. (1999) Comparison of the conformation and the dynamics of saturated and mono-unsaturated hydrocarbon chains of phosphatidylcholines. *Curr Top Biophys.*; **23**: 87–94.
- Murzyn K, Róg T, Jezierski G, Takaoka Y, Pasenkiewicz-Gierula M. (2001) Effects of

- phospholipid unsaturation on the membrane/water interface: a molecular simulation study. *Biophys J.*; **81**: 170–83.
- Pasenkiewicz-Gierula M, Takaoka Y, Miyagawa H, Kitamura K, Kusumi A. (1999) Charge pairing of headgroups in phosphatidylcholine membranes: a molecular dynamics simulation study. *Biophys J.*; **76**: 1228–40.
- Patra M, Karttunen M, Hyvönen MT, Falck E, Linqvist P, Vattulainen I. (2003) Molecular dynamics simulations of lipid bilayers: major artifacts due to truncating electrostatic interactions. *Biophys J.*; **84**: 3646–61.
- Pearlman DA, Case DA, Caldwell JC, Seibel GL, Singh UC, Weiner PK, Kollman PA. (1991) *Amber 4.0*. University of California, San Francisco.
- Róg T, Pasenkiewicz-Gierula M. (2001) Cholesterol effects on the phosphatidylcholine bilayer nonpolar region: A molecular simulation study. *Biophys J.*; **81**: 2190–202.
- Ryckaert JP, Cicotti G, Berendsen HJC. (1977) Numerical integration of the Cartesian equations of motion of a system with constraints: molecular dynamics of n-alkanes. *J Comp Phys.*; **22**: 327–41.
- Sagui C, Darden TA. (1999) Molecular dynamics simulations of biomolecules: long-range electrostatic effects. *Annu Rev Biophys Biomol Struct.*; **28**: 155–79.
- Schreiber H, Steinhauser O. (1992) Cutoff size does strongly influence molecular dynamics results on solvated polypeptides. *Biochemistry.*; **31**: 5856–60.
- Seelig J, Waespe-Sarčević N. (1978) Molecular order in *cis* and *trans* unsaturated phospholipid bilayers. *Biochemistry.*; **17**: 3310–5.
- Shinoda W, Okazaki S. (1998) A Voronoi analysis of lipid area fluctuation in a bilayer. *J Chem Phys.*; **109**: 1517–21.
- Tabony J, Perly B. (1991) Quasielastic neutron scattering measurements of fast local translational diffusion of lipid molecules in phospholipid bilayers. *Biochim Biophys Acta.*; **1063**: 67–72.
- Tieleman DP, Berendsen HJC. (1998) A molecular dynamics study of the pores formed by *Escherichia coli* OmpF porin in a fully hydrated palmitoyloleoylphosphatidylcholine bilayer. *Biophys J.*; **74**: 2786–801.
- Trouard TP, Nevzorov AA, Alam TM, Job CC, Zajicek J, Brown MF. (1999) Influence of cholesterol on dynamics of dimyristoylphosphatidylcholine bilayers as studied by deuterium NMR relaxation. *J Chem Phys.*; **110**: 8802–18.
- York DM, Yang W, Lee H, Darden T, Pedersen LG. (1995) Toward the accurate modeling of DNA: the importance of long-range electrostatics. *J Am Chem Soc.*; **117**: 5001–2.
- 
- \*Róg T, Murzyn K, Gurbel R, Takaoka Y, Kusumi A, Pasenkiewicz-Gierula M. (2003) Effects of phospholipid unsaturation on the bilayer nonpolar region: a molecular simulation study. Submitted to *J Lipid Res*.